www.brjpharmacol.org



### RESEARCH PAPER

# $\beta_3$ -Adrenoceptor agonist stimulation of the Na $^+$ ,K $^+$ -pump in rat skeletal muscle is mediated by $\beta_2$ - rather than $\beta_3$ -adrenoceptors

KT Murphy<sup>1</sup>, H Bundgaard<sup>2</sup> and T Clausen<sup>1</sup>

<sup>1</sup>Institute of Physiology and Biophysics, University of Aarhus, Århus, Denmark and <sup>2</sup>Medical Department B 2142, The Heart Centre, Rigshospitalet, National University Hospital, University of Copenhagen, Copenhagen, Denmark

**Background and purpose:** In cardiac muscle, BRL 37344, a selective  $\beta_3$ -adrenoceptor agonist, activates the Na<sup>+</sup>,K<sup>+</sup>-pump via NO signalling. This study investigated whether BRL 37344 also activates the Na<sup>+</sup>,K<sup>+</sup>-pump via  $\beta_3$ -adrenoceptors in skeletal muscle.

**Experimental approach:** Isolated rat soleus muscles were incubated between 1 and 60 min in buffer. Intracellular Na<sup>+</sup>,K<sup>+</sup> content and Na<sup>+</sup>,K<sup>+</sup>-pump activity were measured using flame photometry and ouabain-suppressible <sup>86</sup>Rb<sup>+</sup> uptake, respectively. Additional muscles were mounted on force transducers and stimulated (60 Hz for 2 s) every 10 min.

**Key results:** BRL 37344 ( $10^{-8}$ - $10^{-5}$  M) induced a concentration- and time-dependent reduction in intracellular Na<sup>+</sup>, and increased ouabain-suppressible <sup>86</sup>Rb<sup>+</sup> uptake by up to 112%. BRL 37344-induced reductions in intracellular Na<sup>+</sup> were blocked by the β<sub>1</sub>/β<sub>2</sub>-adrenoceptor antagonist, nadolol ( $10^{-7}$  M), and the β<sub>2</sub>-adrenoceptor antagonist, ICI 118,551 ( $10^{-7}$ - $10^{-5}$  M), but not by β<sub>3</sub>- or β<sub>1</sub>-adrenoceptor antagonists, SR 59230A ( $10^{-7}$  M) and CGP 20712A ( $10^{-7}$ - $10^{-5}$  M), respectively. Another β<sub>3</sub>-adrenoceptor agonist, CL 316,243, did not alter intracellular Na<sup>+</sup>. BRL 37344-induced reductions in intracellular Na<sup>+</sup> were not blocked by L-NAME, an NOS inhibitor, or ODQ, a guanylyl cyclase inhibitor. The NO donors, SNP and SNAP, did not alter intracellular Na<sup>+</sup>. BRL 37344 rapidly recovered force in muscles depressed by high [K<sup>+</sup>]<sub>o</sub>, an effect that was blocked by nadolol, but not L-NAME.

Conclusions and implications: In rat soleus muscle, the  $\beta_3$ -adrenoceptor agonist BRL 37344 stimulated the Na $^+$ ,K $^+$ -pump via  $\beta_2$ -adrenoceptors. A more selective  $\beta_3$ -adrenoceptor agonist did not affect Na $^+$ , K $^+$  homeostasis in skeletal muscle. NO did not seem to mediate Na $^+$ ,K $^+$ -pump stimulation in skeletal muscle.

British Journal of Pharmacology (2006) 149, 635-646. doi:10.1038/sj.bjp.0706896; published online 3 October 2006

**Keywords:**  $β_3$ -adrenoceptors; Na $^+$ ,K $^+$ -pump; nitric oxide; skeletal muscle;  $β_2$ -adrenoceptors; BRL 37344; Na $^+$ content;  $β_1$ -adrenoceptors; ouabain-suppressible  $^{86}$ Rb $^+$  uptake; K $^+$  content

 $\beta_1$ -adrenoceptors; ouabain-suppressible <sup>86</sup>Rb + uptake; K + content **Abbreviations:** BAT, brown adipose tissue; BRL 37344, 4-(2-[{2-hydroxy-2-(3-chlorophenyl)ethyl}-amino]propyl)-phenoxya-

cetic acid; CGP 20712A, 2-hydroxy-5-(2-[{hydroxy-3-(4-[1-methyl-4-trifluoromethyl-2-imidazolyl]phenoxy)-propyl}amino]ethyoxy) benzamide; CL 316,243, disodium (*R*,*R*)-5-(2-[{2-(3-chlorophenyl)-2-hydroxyethyl}-amino]propyl)-1,3-benzodioxole-2,2-dicarboxylate; ICI 118,551, (—)-1-(2,3-[dihydro-7-methyl-1*H*-inden-4-yl]oxy)-3-([1-methylethyl]-amino)-2-butanol; L-NAME, *N*-nitro-L-arginine methyl ester hydrochloride; NO, nitric oxide; ODQ, 1*H*-[1,2,4]oxadiazolo[4,3,a]quinoaxilin-1-one; PKG, cGMP-dependent protein kinase; SNP, sodium nitroprusside; SNAP, *S*-nitroso-*N*-acetylpenicillamine; SR 59230A, 3-(2-ethylphenoxy)-1-([1*S*]-1,2,3,4-tetrahydronapth-1-ylamino)-2*S*-2-propranol oxalate; TCA, trichloroacetic acid

#### Introduction

In skeletal muscle, the ability to generate repeated muscle contractions depends on the maintenance of membrane excitability, which is compromised by the passive  $\mathrm{Na}^+$  influx and  $\mathrm{K}^+$  efflux associated with each action potential.

Excitability depends on the membrane-bound  $Na^+, K^+$ -ATPase ( $Na^+, K^+$ -pump), which actively transports  $Na^+$  out of the cell, and  $K^+$  back into the cell to restore the transsarcolemmal  $Na^+$  and  $K^+$  gradients. Indeed, during periods of repeated action potentials, the accelerated passive ion fluxes overwhelm the capacity of the  $Na^+, K^+$ -pump, leading to a net  $Na^+$  gain and  $K^+$  loss, a resulting membrane depolarization and a reduction in muscle function (Sejersted and Sjøgaard, 2000).

Correspondence: Dr K Murphy, Institute of Physiology and Biophysics, University of Aarhus, Ole Worms Allé 160, Århus C DK-8000, Denmark. E-mail: km@fi.au.dk

Received 5 July 2006; revised 20 July 2006; accepted 11 August 2006; published online 3 October 2006

In skeletal muscle, the Na<sup>+</sup>,K<sup>+</sup>-pump is acutely stimulated by numerous factors, among which the most potent are the catecholamines and synthetic agonists acting via  $\beta_2$ -adrenoceptors. This leads to activation of adenylate cyclase, increasing cAMP, and resulting in protein kinase A activation (Clausen, 2003). This increases the affinity of the Na<sup>+</sup>,K<sup>+</sup>-pump for intracellular Na<sup>+</sup> (Buchanan *et al.*, 2002), leading to elevated Na<sup>+</sup> efflux and K<sup>+</sup> influx, and mem-

brane hyperpolarization (Clausen and Flatman, 1980).

The  $\beta_3$ -adrenoceptor, or the so-called atypical  $\beta$ -adrenoceptor (Arch and Kaumann, 1993), first became known as a potential anti-obesity target through its effect of increasing energy expenditure and lipolysis in brown adipose tissue (BAT), and therefore the metabolic rate of rats and mice (Arch et al., 1984; Yen et al., 1984). However, similar effects were not found in humans, due to their lack of BAT (reviewed in (Arch and Kaumann (1993)). Whereas expression of the  $\beta_3$ -adrenoceptor is highly abundant in BAT, expression of the  $\beta_3$ -adrenoceptor in skeletal muscle remains controversial. In mammalian skeletal muscle, the  $\beta_3$ -mRNA and protein have been detected in some studies (Evans et al., 1996; Chamberlain et al., 1999), but not in others (Granneman et al., 1991; Thomas and Liggett, 1993; De Matteis et al., 2002). Furthermore, the existence of functional  $\beta_3$ -adrenoceptors in skeletal muscle has been suggested by studies in which selective  $\beta_3$ -adrenoceptor agonists have inhibited proteolysis (Navegantes et al., 2006), and stimulated metabolic oxidation (Board et al., 2000) and glucose utilization (Liu et al., 1996).

Recent evidence in cardiac myocytes suggests that the  $\beta_3$ -adrenoceptor stimulates the Na<sup>+</sup>,K<sup>+</sup>-pump. In mouse cardiac myocytes, the  $\beta_3$ -adrenoceptor agonist, 4-(2-[{2hydroxy-2-(3-chlorophenyl)ethyl}-amino|propyl)-phenoxyacetic acid (BRL 37344), stimulated the endothelial isoform of nitric oxide synthase (NOS), eNOS (Barouch et al., 2002). Stimulation of NOS activates soluble guanylyl cyclase, leading to increased cGMP and activation of cGMP-dependent protein kinase (PKG) (Kobzik et al., 1994). Furthermore, in rabbit cardiac myocytes, the nitric oxide (NO) donor, sodium nitroprusside (SNP), stimulated the Na<sup>+</sup>,K<sup>+</sup>-pumpmediated current, an effect that was abolished by inhibition of both guanylyl cyclase and PKG (William et al., 2005). In the same preparation, BRL 37344 stimulated the Na<sup>+</sup>,K<sup>+</sup>pump-mediated current, an effect that was abolished by inhibition of NOS (Bundgaard et al., 2006a). Taken together, these results indicate that in cardiac myocytes,  $\beta_3$ -adrenoceptor agonists stimulate the Na+,K+-pump and that this effect is mediated via activation of NOS and signalling by NO. As this signalling is likely to be restricted to sarcolemmal microdomains with a large content of Na<sup>+</sup>,K<sup>+</sup>-pumps (Liu et al., 2003), NO-induced stimulation of the Na<sup>+</sup>, K<sup>+</sup>-pump may be more selective and rapid than  $\beta_2$ -adrenoceptor stimulation mediated by cAMP. This would be important in skeletal muscle, since a rapid stimulation of the Na<sup>+</sup>,K<sup>+</sup>pump would delay the loss in membrane excitability, and hence, the decline of muscle function associated with repeated muscle contractions. This would be of further physiological interest because the  $\beta$ -adrenergic system in skeletal muscle would not only involve effects mediated by cAMP, but possibly also cGMP. However, it is unknown

whether  $\beta_3$ -adrenoceptors and/or NO stimulate the Na  $^+$ ,K  $^+$ -pump in skeletal muscle.

The primary aim of this study was therefore to investigate the effect of the  $\beta_3$ -adrenoceptor agonist, BRL 37344, on the Na $^+$ ,K $^+$ -pump in isolated rat skeletal muscle. This would also yield information regarding the possible existence of the  $\beta_3$ -adrenoceptor in skeletal muscle. The second aim was to investigate whether Na $^+$ ,K $^+$ -pump stimulation in skeletal muscle can be mediated by an NO signalling system.

Here, we show that the  $\beta_3$ -adrenoceptor agonist, BRL 37344, stimulates the Na<sup>+</sup>,K<sup>+</sup>-pump but that this occurs via the  $\beta_2$ -, rather than the  $\beta_3$ -adrenoceptors. Results with the more selective  $\beta_3$ -adrenoceptor agonist, disodium (R,R)-5-(2-[{2-(3-chlorophenyl)-2-hydroxyethyl}-amino]propyl)-1,3-benzodioxole-2,2-dicarboxylate (CL 316,243) (Dolan *et al.*, 1994), suggest that  $\beta_3$ -adrenoceptors expressed in skeletal muscle are not involved in Na<sup>+</sup> and K<sup>+</sup> homeostasis. Furthermore, NO does not appear to mediate Na<sup>+</sup>,K<sup>+</sup>-pump stimulation in rat skeletal muscle.

#### Methods

#### Animals and preparation of muscles

Experiments were carried out using 4-week-old Wistar rats, weighing approximately 60–70 g. The animals were fed *ad libitum* and were maintained in a temperature-controlled environment (21°C) with constant day length (12h). The animals were killed by cervical dislocation, followed by decapitation, with intact soleus muscles, a predominantly slow-twitch fibre muscle (weighing 20–30 mg), dissected out as previously described (Nielsen and Clausen, 1996). All handling and use of animals complied with Danish animal welfare regulations.

Muscles were equilibrated for 30 min at 30 °C in standard Krebs–Ringer bicarbonate buffer (KR) (pH 7.4), containing the following (in mm): 122.1 NaCl, 25.1 NaHCO $_3$ , 2.8 KCl, 1.2 KH $_2$ PO $_4$ , 1.2 MgSO $_4$ , 1.3 CaCl $_2$  and 5.0 D-glucose, and were bubbled continuously with a mixture of 95% O $_2$  and 5% CO $_2$ . In buffer with 11.0 mM K $^+$ , an equivalent amount of Na $^+$  was omitted to maintain iso-osmolarity.

#### Incubation experiments

For each of the following interventions, muscles were placed in polyethylene baskets and following equilibration for 30 min in standard KR, were incubated at 30°C in the appropriate buffers for the indicated durations (final volume 2 ml). Control muscles were incubated for durations matching their respective experimental muscles in standard KR.

#### *Effect of* $\beta_3$ -adrenoceptor agonists

The majority of experiments involved incubating muscles between 1 and 60 min in KR containing the  $\beta_3$ -adrenoceptor agonist, BRL 37344 ( $10^{-10}$ – $10^{-5}$  M) (Langin *et al.*, 1991). To further test the effect of  $\beta_3$ -adrenoceptor activation on Na<sup>+</sup> homeostasis, some experiments involved incubating muscles for 30 min in KR containing the more selective

 $\beta_3$ -adrenoceptor agonist, CL 316,243 ( $10^{-7}$  M,  $10^{-5}$  M) (Dolan *et al.*, 1994).

#### Effect of β-adrenoceptor antagonists

To investigate whether the effect of BRL 37344 on intracellular Na $^+$  content was mediated via the  $\beta_3$ -,  $\beta_1/\beta_2$ -,  $\beta_2$ - or  $\beta_1$ -adrenoceptors, muscles were preincubated for 15 min in KR containing the  $\beta_3$ -adrenoceptor antagonist, 3-(2-ethylphenoxy)-1-([1S]-1,2,3,4-tetrahydronapth-1-ylamino)-2S-2-propanol oxalate (SR 59230A); ( $10^{-7}$  M,  $10^{-5}$  M) (Nisoli *et al.*, 1996), the  $\beta_1/\beta_2$ -adrenoceptor antagonist, nadolol ( $10^{-7}$  M) (Bond and Clarke, 1988), the selective  $\beta_2$ -adrenoceptor antagonist, (–)-1-(2,3-[dihydro-7-methyl-1*H*-inden-4-yl]oxy)-3-([1-methylethyl]-amino)-2-butanol (ICI 118,551); ( $10^{-7}$  M,  $10^{-5}$  M) (O'Donnell and Wanstall, 1980) or the selective  $\beta_1$ -adrenoceptor antagonist, 2-hydroxy-5-(2-[{hydroxy-3-(4-[1-methyl-4-trifluoromethyl-2-imidazolyl]phenoxy)propyl} amino]ethyoxy) benzamide (CGP 20712A); ( $10^{-7}$  M,  $10^{-5}$  M) (Dooley *et al.*, 1986), respectively.

#### Selectivity of β-adrenoceptor antagonists

To investigate the selectivity of SR 59230A (for  $\beta_3$ -adrenoceptors), nadolol (for  $\beta_1/\beta_2$ - adrenoceptors), ICI 118,551 (for  $\beta_2$ -adrenoceptors) and CGP 20712A (for  $\beta_1$ -adrenoceptors), muscles were preincubated for 15 min in KR containing SR 59230A ( $10^{-7}$  M,  $10^{-5}$  M), nadolol ( $10^{-7}$  M), ICI 118,551 ( $10^{-5}$  M) or CGP 20712A ( $10^{-5}$  M), and were then incubated for 30 min in KR containing the  $\beta_2$ -adrenoceptor agonist, salbutamol ( $10^{-7}$  M) (Baker, 2005) with or without SR 59230A ( $10^{-7}$  M,  $10^{-5}$  M), nadolol ( $10^{-7}$  M), ICI 118,551 ( $10^{-5}$  M) or CGP 20712A ( $10^{-5}$  M).

#### Effect of NO inhibitors and donors

Inhibition of NOS was induced by preincubating muscles for 60 min in KR containing *N*-nitro-L-arginine methyl ester hydrochloride (L-NAME;  $10^{-5}$  M,  $10^{-3}$  M), an inhibitor of NOS (Rees *et al.*, 1990). 1*H*-[1,2,4]oxadiazolo[4,3,a]quinoaxilin-1-one (ODQ) ( $10^{-5}$  M) was used to inhibit NO-sensitive guanylyl cyclase (Garthwaite *et al.*, 1995).

Muscles were exposed to NO by incubating muscles for 30 min in KR containing SNP ( $10^{-5}$  M,  $10^{-4}$  M) (Young and Leighton, 1998a) or for 60 min in KR containing *S*-nitroso-*N*-acetylpenicillamine (SNAP;  $10^{-5}$ – $10^{-3}$  M), a nitrosothiol derivative (Holm *et al.*, 1998).

## Effect of noradrenaline – mediated via the $\beta_3$ -, $\beta_2$ - or $\beta_1$ -adrenoceptors

To investigate whether the reduction in intracellular Na $^+$  content induced by noradrenaline was mediated via the  $\beta_3$ -,  $\beta_2$ - or  $\beta_1$ -adrenoceptors, muscles were preincubated for 15 min in KR containing SR 59230A ( $10^{-7}$  M), nadolol ( $10^{-6}$  M), ICI 118,551 ( $10^{-7}$  M) or CGP 20712A ( $10^{-7}$  M), and were then incubated for 30 min in KR containing noradrenaline ( $10^{-5}$  M) with or without SR 59230A ( $10^{-7}$  M), nadolol ( $10^{-6}$  M), ICI 118,551 ( $10^{-7}$  M) or CGP 20712A ( $10^{-7}$  M).

Measurement of intracellular  $Na^+$  and  $K^+$  contents

Following incubation, muscles were immediately transferred to ice-cold Na $^+$ -free Tris-sucrose buffer and underwent a  $4\times15$  min washout to remove extracellular Na $^+$ . Following washout, muscles were blotted, tendons cut off, muscle wet weight determined, and soaked overnight in 0.3 M trichloroacetic acid (TCA) to give complete extraction of ions from the tissue (Clausen *et al.*, 1993). The Na $^+$  and K $^+$  contents in the TCA extract was measured by flame photometry (FLM3, Radiometer, Copenhagen, Denmark) with lithium as internal standard. Values for Na $^+$  content were then multiplied by 1.46 to correct for the loss of intracellular Na $^+$  during the ice-cold washout (Everts and Clausen, 1992). In contrast, the loss of K $^+$  during the washout was minimal (Everts and Clausen, 1992).

Measurement of <sup>86</sup>Rb<sup>+</sup> uptake rate and Na<sup>+</sup>,K<sup>+</sup>-pump activity  $^{86}\mathrm{Rb^+}$  has previously been shown to be a reliable tracer for determination of Na<sup>+</sup>,K<sup>+</sup>-pump-mediated K<sup>+</sup> transport (Clausen et al., 1987). To investigate the time course of the effects of BRL 37344 on total  $^{86}\mathrm{Rb}^+$  uptake, following equilibration, muscles were incubated between 1 and 20 min in buffer containing  $^{86}\text{Rb}^+$  (0.1  $\mu\text{Ci ml}^{-1}$ ) without or with  $10^{-5}$  M BRL 37344. In the next series of experiments, we investigated the effect of BRL 37344 on Na<sup>+</sup>,K<sup>+</sup>-pump activity, by measuring ouabain-sensitive <sup>86</sup>Rb<sup>+</sup> uptake. Following equilibration, muscles were preincubated for 15 min without or with  $10^{-3}$  M ouabain, followed by a further 5 min incubation in KR containing  $^{86}\text{Rb}^+$  (0.1  $\mu\text{Ci ml}^{-1}$ ) without or with  $10^{-3}$  M ouabain and/or  $10^{-5}$  M BRL 37344. All muscles then underwent a  $4 \times 15$  min washout in ice-cold Na<sup>+</sup>-free Tris-sucrose buffer to remove extracellular <sup>86</sup>Rb<sup>+</sup> and Na<sup>+</sup>. Following washout, muscles were blotted, tendons cut off, muscle wet weight determined, and soaked overnight in 2 ml 0.3 M TCA in 4 ml counting vials. Muscles were then taken for counting of 86Rb+ activity by Cerenkov radiation in a  $\beta$ -counter. The amount of  $^{86}\text{Rb}^+$  activity retained after the washout was calculated and expressed as the relative uptake of the <sup>86</sup>Rb<sup>+</sup> activity from the incubation medium by the muscle. The K<sup>+</sup> uptake was then calculated by converting the relative uptake of <sup>86</sup>Rb<sup>+</sup> to K<sup>+</sup> using the concentration of K<sup>+</sup> in the incubation medium (for details see (Buchanan et al., 2002)). Previous studies have reported that the loss of 86Rb+ during the washout was minimal (Buchanan et al., 2002), and therefore no correction was made for <sup>86</sup>Rb<sup>+</sup> uptake values.

#### Measurement of <sup>22</sup>Na<sup>+</sup> influx

The effect of BRL 37344 on the Na $^+$  influx was determined by measuring the initial rate of  $^{22}$ Na $^+$  influx, as described elsewhere (Clausen and Kohn, 1977). Following equilibration, muscles were preincubated for 15 min in KR without or with  $10^{-3}$  M ouabain. Muscles were then incubated for 2 min in KR containing  $^{22}$ Na $^+$  (0.5  $\mu$ Ci ml $^{-1}$ ) without or with  $10^{-3}$  M ouabain and without or with  $10^{-5}$  M BRL 37344. All muscles then underwent a  $4 \times 15$  min washout in ice-cold Na $^+$ -free Tris-sucrose buffer to remove all extracellular  $^{22}$ Na $^+$ . Following washout, muscles were blotted, tendons

cut off, muscle wet weight determined, and the activity of the  $^{22}\mathrm{Na}^{+}$  retained in the muscles was determined by  $\gamma$ -counting. After correction for the loss of intracellular  $^{22}\mathrm{Na}^{+}$  during the washout, the Na $^{+}$  influx was calculated from the specific activity of  $^{22}\mathrm{Na}^{+}$  in the incubation buffer (Clausen and Kohn, 1977).

#### Measurement of force

Intact muscles were mounted at optimal force generating length on electrodes for isometric contractions and equilibrated for 30 min in KR at 30°C. Muscles were then exposed to field stimulation across the central region through platinum electrodes, using 2 s trains of 0.2 ms 12 V pulses at 60 Hz every 10 min. Force was measured using force displacement transducers and recorded with a chart recorder and/or digitally on a computer. The mean absolute force produced under control conditions was  $0.37 \pm 0.02 \, \mathrm{N}$  (n = 24), with results expressed as a percentage of the control force produced in standard KR.

#### Chemicals and isotopes

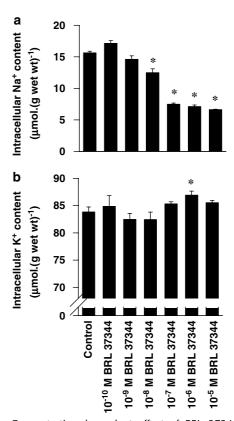
All chemicals were of analytical grade. BRL 37344, ouabain, nadolol, SR 59230A, salbutamol, CL 316,243, CGP 20712A, ICI 118,551, L-NAME, ODQ , SNP and SNAP were purchased from Sigma Chemicals (St Louis, MO, USA).  $^{86}{\rm Rb}^+$  (0.1  $\mu{\rm Ci~ml}^{-1}$  buffer) and  $^{22}{\rm Na}^+$  (0.5  $\mu{\rm Ci~ml}^{-1}$  buffer) was from Amersham International (Aylesbury, Buckinghamshire, UK).

#### Statistical analysis

All data are presented as either mean  $\pm$  s.e.m. or for the sake of clarity as mean with s.e.m. The statistical differences between two groups was analysed using a nonpaired-samples Student's *t*-test. The statistical difference between three or more groups was analysed using a one-way ANOVA. Differences were located with a Student–Newman–Keuls *post hoc* test. Significance was accepted at P < 0.05.

#### Results

Concentration- and time-dependent effects of BRL 37344 The effect of six different concentrations of BRL 37344  $(10^{-10}$ – $10^{-5}$  M, all 60 min) on the intracellular Na<sup>+</sup> and K<sup>+</sup> contents was initially investigated (Figure 1). There was no effect of BRL 37344 on intracellular Na+ content at concentrations of  $10^{-10}$  M and  $10^{-9}$  M (Figure 1a). However, intracellular Na + content was significantly reduced with concentrations of BRL 37344 exceeding 10<sup>-9</sup> M, with the largest reduction (56%) occurring with 10<sup>-5</sup> M BRL 37344 (Figure 1a). In contrast, the only significant effect of BRL 37344 on intracellular K<sup>+</sup> content occurred at a concentration of  $10^{-6}$  M, where intracellular K<sup>+</sup> content was 5% higher than in control muscles (P < 0.01, Figure 1b). In the presence of ouabain at a concentration  $(10^{-3} \,\mathrm{M})$  sufficient to block the Na+,K+-pumps, BRL 37344 (10-5 M) produced no significant change in Na<sup>+</sup> (+ouabain,  $29.5\pm1.4$ ; ouabain + BRL 37344,  $27.9 \pm 0.8 \,\mu\text{mol}(\text{g wet wt})^{-1}$ , n = 4) or



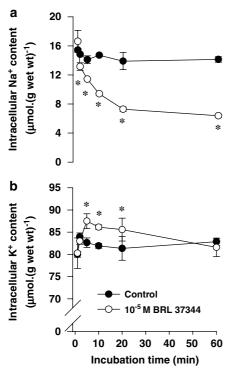
**Figure 1** Concentration-dependent effect of BRL 37344 on (a) intracellular Na $^+$  and (b) K $^+$  contents in rat soleus muscle. Muscles were placed in polyethylene baskets, equilibrated for 30 min in standard KR and then incubated for 60 min without or with  $10^{-10}$ –  $10^{-5}$  M BRL 37344. Muscles were then washed for  $4 \times 15$  min in iccold Na $^+$ -free Tris-sucrose buffer, blotted, tendons removed, weighed and taken for flame photometric analysis of Na $^+$  and K $^+$  content. Data are means with s.e.m.; n=4–11. \*P<0.01 vs control.

 $\rm K^+$  contents (+ouabain, 70.5±0.3; ouabain+BRL 37344, 69.8±1.6 μmol(g wet wt)<sup>-1</sup>, n=4). Moreover, in comparison to the control values given in Figure 1, ouabain alone produced a large increase in intracellular Na<sup>+</sup> content and a large reduction in intracellular K<sup>+</sup> content.

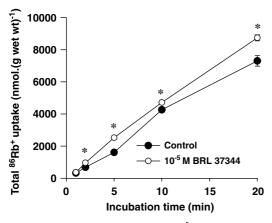
Figure 2 shows the time course of the effect of BRL 37344  $(10^{-5}\,\mathrm{M})$  on intracellular Na  $^+$  and K  $^+$  content. The reduction in intracellular Na  $^+$  content was significant as early as 2 min after the onset of incubation with BRL 37344 (11%) and the decline continued until after 20–60 min when the intracellular Na  $^+$  content approached a plateau around 50% lower than controls (Figure 2a). The elevation in intracellular K  $^+$  content with BRL 37344 was significantly different from controls at 5 (6%), 10 (5%) and 20 min after the onset of incubation (5%), Figure 2b).

BRL 37344  $(10^{-5} \text{ M})$  also increased total  $^{86}\text{Rb}^+$  uptake, by 42, 57, 11 and 20% at 2, 5, 10 and 20 min following the onset of incubation, respectively (Figure 3). This indicates an early stimulation of the Na $^+$ ,K $^+$ -pump.

Effect of BRL 37344 on  $Na^+$ , $K^+$ -pump activity Ouabain (10<sup>-3</sup> M) decreased <sup>86</sup>Rb<sup>+</sup> uptake from 394±23 to  $177\pm7$  nmol(g wet wt)<sup>-1</sup> min<sup>-1</sup> (n=4). The difference, 216 $\pm$ 26 nmol(g wet wt)<sup>-1</sup>, is a measure of Na<sup>+</sup>,K<sup>+</sup>-pump activity. Incubation with  $10^{-5}$  M BRL 37344 (5 min) increased this ouabain-sensitive <sup>86</sup>Rb<sup>+</sup> uptake to  $458\pm12$  nmol (g wet wt)<sup>-1</sup> min<sup>-1</sup> (n=4). In the presence of ouabain, there was no effect of BRL 37344 on <sup>86</sup>Rb<sup>+</sup> uptake (+ouabain,  $177\pm7$ ; ouabain + BRL 37344,  $165\pm9$  nmol(g wet wt)<sup>-1</sup> min<sup>-1</sup>, n=4). These results indicate that the increase in total <sup>86</sup>Rb<sup>+</sup> uptake with BRL 37344 (Figure 3) was due to increased Na<sup>+</sup>,K<sup>+</sup>-pump activity.



**Figure 2** Time course of the effects of  $10^{-5}$  M BRL 37344 on (a) intracellular Na<sup>+</sup> and (b) K<sup>+</sup> contents in rat soleus muscle. Muscles were incubated for 1, 2, 5, 10, 20 or 60 min without or with  $10^{-5}$  M BRL 37344. Experimental conditions as described in Figure 1. Data are means  $\pm$  s.e.m.; n=4-8. \*P<0.05 vs control.



**Figure 3** Time course of the effects of  $10^{-5}$  M BRL 37344 on total  $^{86}$ Rb  $^+$  uptake in rat soleus muscle. Muscles were transferred into KR containing  $^{86}$ Rb  $^+$  (0.1  $\mu$ Ci ml $^{-1}$ ) and incubated for 1, 2, 5, 10 or 20 min without or with  $10^{-5}$  M BRL 37344. Experimental conditions as described in Figure 1. Data are means  $\pm$  s.e.m.; n = 4. \*p < 0.02 vs control.

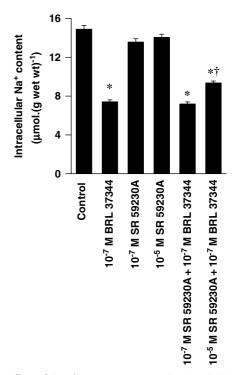
Effect of BRL 37344 on Na+ influx

Incubation with  $10^{-5}$  M BRL 37344 did not significantly attenuate  $^{22}$ Na $^+$  influx measured over a 2 min period neither in the absence (control,  $598\pm22$ ; +BRL 37344,  $564\pm18$  nmol g $^{-1}$  min $^{-1}$ , n=4-5) nor in the presence of  $10^{-3}$  M ouabain (+ ouabain,  $784\pm42$ ; ouabain + BRL 37344,  $705\pm8$  nmol g $^{-1}$  min $^{-1}$ , n=5).

Effect of  $\beta$ -adrenoceptor antagonists on the BRL 37344-induced reduction in intracellular Na  $^+$  content

The β<sub>3</sub>-adrenoceptor antagonist, SR 59230A. At a concentration of  $10^{-7}$  M, the β<sub>3</sub>-adrenoceptor antagonist, SR 59230A, had no effect on the reduction in intracellular Na<sup>+</sup> content induced by  $10^{-7}$  M BRL 37344 (Figure 4). However at a concentration of  $10^{-5}$  M, SR 59230A attenuated the reduction in intracellular Na<sup>+</sup> content induced by  $10^{-7}$  M BRL 37344, by 13% (Figure 4).

The  $\beta_1/\beta_2$ -adrenoceptor antagonist, nadolol and the  $\beta_2$ -adrenoceptor antagonist, ICI 118,551. Preincubation with the  $\beta_1/\beta_2$ -adrenoceptor antagonist, nadolol  $(10^{-7}\,\mathrm{M})$  and with the selective  $\beta_2$ -adrenoceptor antagonist, ICI 118,551  $(10^{-5}\,\mathrm{M})$ , completely suppressed the reduction in intracellular Na<sup>+</sup> content induced by  $10^{-7}\,\mathrm{M}$  BRL 37344 (Figure 5). At the lower concentration of  $10^{-7}\,\mathrm{M}$ , preincubation with ICI 118,551 partially suppressed the reduction in intracellular Na<sup>+</sup> content induced by BRL 37344 (control,  $15.7\pm0.5$ ;



**Figure 4** Effect of  $\beta_3$ -adrenoceptor antagonist, SR 59230A, on BRL 37344-induced change in intracellular Na $^+$  content in rat soleus muscle. Muscles were preincubated for 15 min without or with  $10^{-7}$  M or  $10^{-5}$  M SR 59230A, incubated for 30 min without or with the indicated additions, and were then treated as described in Figure 1. Data are means with s.e.m.; n=4. \*P<0.01 vs control,  $^{\dagger}P<0.001$  vs  $10^{-7}$  M BRL 37344.

+ BRL 37344, 9.2  $\pm$  0.3; ICI 118,551 + BRL 37344, 12.4  $\pm$  0.5  $\mu$ mol(g wet wt)<sup>-1</sup>, n = 4–12).

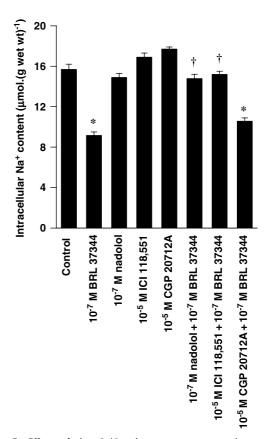
The  $\beta_1$ -adrenoceptor antagonist, CGP 20712A. Preincubation with the selective  $\beta_1$ -adrenoceptor antagonist, CGP 20712A ( $10^{-5}\,\mathrm{M}$ ), had no effect on the reduction in intracellular Na $^+$  content induced by  $10^{-7}\,\mathrm{M}$  BRL 37344 (Figure 5). Similar effects were found with  $10^{-7}\,\mathrm{M}$  CGP 20712A (control,  $15.7 \pm 0.5$ ; +BRL 37344,  $9.2 \pm 0.3$ ; CGP 20712A +BRL 37344,  $10.0 \pm 0.7\,\mu\mathrm{mol}(\mathrm{g}\,\mathrm{wet}\,\mathrm{wt})^{-1}$ , n = 4–12).

Selectivity of SR 59230A, nadolol, ICI 118,551 and CGP 20712A The effects of preincubation with SR 59230A ( $10^{-7}$  M,  $10^{-5}$  M), nadolol ( $10^{-7}$  M), ICI 118,551 ( $10^{-5}$  M) and CGP 20712A ( $10^{-5}$  M) on the reduction in intracellular Na<sup>+</sup> content induced by the  $\beta_2$ -adrenoceptor agonist, salbutamol ( $10^{-7}$  M), was used to assess the selectivity of these antagonists (Table 1).

Salbutamol induced a 47% reduction in intracellular Na $^+$  content, that was not blocked by preincubation with  $10^{-7}\,\rm M$  SR 59230A, but was attenuated to a 38% reduction compared to controls by preincubation with  $10^{-5}\,\rm M$  SR 59230A (Table 1). The reduction in intracellular Na $^+$  content induced by salbutamol was blocked by preincubation with nadolol and ICI 118,551 but not CGP 20712A (Table 1).

## Effect of the $\beta_3$ -adrenoceptor agonist, CL 316,243, on intracellular Na $^+$ content

To further investigate whether  $\beta_3$ -adrenoceptor agonists were involved in Na<sup>+</sup> homeostasis in skeletal muscle, the effects of the more selective  $\beta_3$ -adrenoceptor agonist, CL 316,243, on intracellular Na<sup>+</sup> content was examined. There was no effect of CL 316,243 on intracellular Na<sup>+</sup> content at any of the concentrations tested (control,  $14.8 \pm 0.6$ ;  $+10^{-7}$  M CL 316,243,  $14.1 \pm 0.3$ ;  $+10^{-5}$  M CL 316,243,  $14.2 \pm 0.5$   $\mu$ mol(g wet wt)<sup>-1</sup>, all n = 6-7).



**Figure 5** Effect of the  $β_1/β_2$ -adrenoceptor antagonist, nadolol,  $β_2$ -adrenoceptor antagonist, ICI 118,551, and  $β_1$ -adrenoceptor antagonist, CGP 20712A, on BRL 37344-induced change in intracellular Na $^+$  content in rat soleus muscle. Muscles were preincubated for 15 min without or with  $10^{-7}$  M nadolol,  $10^{-5}$  ICI 118,551 or  $10^{-5}$  M CGP 20712A, incubated for 30 min without or with the indicated additions and were then treated as described in Figure 1. Data are means with s.e.m.; n=4–12. \*P<0.001 vs control,  $^†P$ <0.001 vs  $10^{-7}$  M BRL 37344.

Table 1 Effects of SR 59230A, nadolol, ICI 118,551 and CGP 20712A on the salbutamol-induced change in intracellular Na<sup>+</sup> content in rat soleus muscle

Preincubation (15 min)	Incubation (30 min)	Intracellular Na $^+$ content ( $\mu$ mol(g wet wt) $^{-1}$ )
No additions	No additions	15.2±0.7
No additions	10 <sup>-7</sup> м Salbutamol	8.0±0.3*
10 <sup>-7</sup> м SR 59230A	10 <sup>-7</sup> м SR 59230А	$14.8 \pm 0.8$
10 <sup>-5</sup> м SR 59230A	10 <sup>-5</sup> м SR 59230A	$14.1 \pm 0.6$
10 <sup>-7</sup> м Nadolol	10 <sup>-7</sup> м Nadolol	$15.4 \pm 1.0$
10 <sup>-5</sup> м ICI 118,551	10 <sup>-5</sup> м ICI 118,551	$16.8 \pm 0.3$
10 <sup>-5</sup> м СGР 20712A	10 <sup>-5</sup> м СGР 20712А	$16.8 \pm 0.4$
10 <sup>-7</sup> м SR 59230A	$10^{-7}$ M SR 59230A $+ 10^{-7}$ M salbutamol	7.7±0.4*
10 <sup>-5</sup> м SR 59230A	$10^{-5}$ M SR 59230A $+ 10^{-7}$ M salbutamol	$9.4 \pm 0.4^{*,\dagger}$
10 <sup>-7</sup> м Nadolol	$10^{-7}$ M Nadolol $+ 10^{-7}$ M salbutamol	$13.2 \pm 0.6^{\dagger}$
10 <sup>-5</sup> м ICI 118,551	$10^{-5}$ M ICI 118,551 $+ 10^{-7}$ M salbutamol	$17.0\pm0.3^{\dagger}$
10 <sup>-5</sup> м СGР 20712A	$10^{-5}\text{M}$ CGP $20712A+10^{-7}\text{M}$ salbutamol	$8.8 \pm 0.2*$

Abbreviations: CGP 20712A, 2-hydroxy-5-(2-[{hydroxy-3-(4-[1-methyl-4-trifluoromethyl-2-imidazolyl]phenoxy)propyl}amino]ethyoxy) benzamide; ICI 118,551, (–)-1-(2,3-[dihydro-7-methyl-1*H*-inden-4-yl]oxy)-3-([1-methylethyl]-amino)-2-butanol; KR, Krebs–Ringer bicarbonate buffer; SR 59230A, 3-(2-ethylphenoxy)-1-([1*S*]-1,2,3,4-tetrahydronapth-1-ylamino)-2*S*-2-propranol oxalate.

Muscles were placed in polyethylene baskets, equilibrated for 30 min in standard KR and preincubated for 15 min without or with  $10^{-7}$  M SR 59230A,  $10^{-5}$  M SR 59230A,  $10^{-7}$  M nadolol,  $10^{-5}$  M ICI 118,551 or  $10^{-5}$  M CGP 20712A. Muscles were then incubated for 30 min without or with the indicated additions, washed for  $4 \times 15$  min in ice-cold Na<sup>+</sup>-free Tris-sucrose buffer, blotted, tendons removed, weighed and taken for analysis of Na<sup>+</sup> content. Data are means  $\pm$  s.e.m., n = 4–10. \*P < 0.001 vs control,  $^{\dagger}P < 0.05$  vs  $10^{-7}$  M salbutamol.

Table 2 Effect of the NOS inhibitors, L-NAME and ODQ, on the BRL 37344-induced change in intracellular Na+ content in rat soleus muscle

Preincubation (60 min)	Incubation (5 min)	Intracellular Na $^+$ content ( $\mu$ mol(g wet wt) $^{-1}$ )
No additions	No additions	14.8±0.3
No additions	$10^{-7}$ M BRL 37344	12.9±0.3*
No additions	$10^{-5}$ M BRL 37344	11.4±0.1*
10 <sup>-5</sup> м L-NAME	$10^{-5}$ M L-NAME	$14.4\pm0.3$
$10^{-3}$ M L-NAME	$10^{-3}$ M L-NAME	$14.8 \pm 0.6$
10 <sup>-5</sup> м ODQ	$10^{-5}$ M ODQ	$14.5 \pm 0.3$
10 <sup>-5</sup> м L-NAME	$10^{-5}$ M L-NAME $+ 10^{-7}$ M BRL 37344	12.8 ± 0.2*
10 <sup>-5</sup> м L-NAME	$10^{-5}$ m L-NAME $+10^{-5}$ m BRL 37344	$10.6\pm0.4*$
$10^{-3}$ M L-NAME	$10^{-3}$ M L-NAME $+10^{-5}$ M BRL 37344	11.3±0.2*
10 <sup>-5</sup> м ODQ	$10^{-5}$ M ODQ $+ 10^{-7}$ M BRL 37344	13.0±0.4*

Abbreviations: BRL 37344, 4-(2-[{2-hydroxy-2-(3-chlorophenyl)ethyl}-amino]propyl)-phenoxyacetic acid; L-NAME, *N*-nitro-L-arginine methyl ester hydrochloride; NOS, nitric oxide synthase; ODQ, 1*H*-[1,2,4]oxadiazolo[4,3,a]quinoaxilin-1-one.

Muscles were preincubated for 60 min without or with  $10^{-5}$  M or  $10^{-3}$  M L-NAME, or  $10^{-5}$  M ODQ, were incubated for 5 min without or with the indicated additions, and then treated as described in Table 1. Data are means  $\pm$  s.e.m., n = 4–8.

Table 3 Effect of NO donors, SNP and SNAP, on intracellular  ${\rm Na}^+$  content in rat soleus muscle

Incubation (30 or 60 min)	Intracellular Na <sup>+</sup> content (µmol(g wet wt) <sup>-1</sup> )
Control	15.9+0.5
10 <sup>-5</sup> м SNP	14.6+0.4
10 <sup>-4</sup> м SNP	$14.7 \pm 0.1$
10 <sup>-5</sup> м SNAP	$17.0\pm0.5$
10 <sup>-4</sup> м SNAP	$15.3 \pm 0.3$
$10^{-3}$ M SNAP	$16.1 \pm 0.5$

Abbreviations: NO, nitric oxide; NS, nonsignificant; SNP, sodium nitroprusside; SNAP, S-nitroso-*N*-acetylpenicillamine.

Muscles were incubated for 30 min without or with  $10^{-5}\,\mathrm{M}$  or  $10^{-4}\,\mathrm{M}$  SNP or for 60 min without or with  $10^{-5}\,\mathrm{M}$ ,  $10^{-4}\,\mathrm{M}$  or  $10^{-3}\,\mathrm{M}$  SNAP. Muscles were then treated as described in Table 1. Data are means  $\pm$  s.e.m., n = 3–4. NS.

Effect of NOS inhibitors and NO donors on the BRL 37344-induced reduction in intracellular  ${\rm Na}^+$  content

Muscles were preincubated with the NOS inhibitor, L-NAME  $(10^{-5}\,\text{M},\ 10^{-3}\,\text{M})$  or with the guanylyl cyclase inhibitor, ODQ  $(10^{-5}\,\text{M})$ , to investigate whether the Na<sup>+</sup>,K<sup>+</sup>-pump stimulation induced by BRL 37344, leading to a reduction in intracellular Na<sup>+</sup> content, was mediated via increased NO signalling. Neither L-NAME nor ODQ caused any significant change in intracellular Na<sup>+</sup> content or in the reduction in intracellular Na<sup>+</sup> content induced by 5 min incubation with BRL 37344  $(10^{-7}\,\text{M},\ 10^{-5}\,\text{M},\ Table\ 2)$ . There was also no effect of L-NAME  $(10^{-3}\,\text{M})$  on the reduction in intracellular Na<sup>+</sup> content induced by 5 min incubation with  $10^{-5}\,\text{M}$  salbutamol (control,  $15.5\pm0.2$ ; + salbutamol,  $10.3\pm0.1$ ; + L-NAME,  $14.9\pm0.3$ ; L-NAME + salbutamol,  $10.2\pm0.2\,\mu\text{mol}(\text{g\,wet\,wt})^{-1},\ n=4)$ .

If increased NO were involved in Na $^+$ ,K $^+$ -pump stimulation in skeletal muscle, the NO donors SNP and SNAP would be expected to reduce intracellular Na $^+$  content. However, there was no significant effect of either SNP (30 min) or SNAP (60 min) on intracellular Na $^+$  content at any of the concentrations tested (SNP,  $10^{-5}$ – $10^{-4}$  M; SNAP,  $10^{-5}$ – $10^{-3}$  M, Table 3).

Effect of nadolol or L-NAME on the BRL 37344-induced force recovery in muscles depressed by high  $[K^+]_0$ 

In human skeletal muscle, intense exercise has been shown to increase the interstitial  $[K^+]$  to as high as  $11-12\,\text{mM}$ 

(Nordsborg *et al.*, 2003). Under these conditions, Na $^+$ ,K $^+$ -pump stimulation is essential to restore the *trans*-sarco-lemmal K $^+$  gradients to protect membrane excitability. It was therefore investigated whether BRL 37344 could induce force recovery in muscles depressed by high extracellular [K $^+$ ] ([K $^+$ ] $_{\rm o}$ ) (11 mM), and whether any force recovery could be blocked by preincubation with nadolol or L-NAME (Figure 6).

Incubation of muscles at  $11\,\mathrm{mM}$  K $^+$  reduced force to  $16\pm5\%$  of initial force (Figure 6a). In these muscles, BRL 37344 produced a rapid force recovery, with force returning to  $70\pm12$  and  $92\pm1\%$  of initial force, at only 10 min following the addition of  $10^{-7}\,\mathrm{M}$  and  $10^{-5}\,\mathrm{M}$  BRL 37344, respectively (Figure 6a). Very similar force recoveries were also seen with  $10^{-7}\,\mathrm{M}$  and  $10^{-5}\,\mathrm{M}$  salbutamol (data not shown). The force recovery observed with  $10^{-5}\,\mathrm{M}$  BRL 37344 was completely blocked by preincubation with  $10^{-5}\,\mathrm{M}$  nadolol (Figure 6a), but was not affected by preincubation with  $10^{-5}\,\mathrm{M}$  L-NAME (Figure 6b).

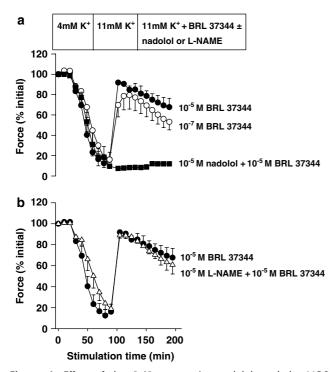
Effect of SR 59230A, nadolol, ICI 118,551 and CGP 20712A on the noradrenaline-induced reduction in intracellular Na<sup>+</sup> content

As shown in Table 4,  $10^{-5}$  M noradrenaline induced a 52% reduction in intracellular Na<sup>+</sup> content, that was not altered by preincubation with either  $10^{-7}$  M SR 59230A or  $10^{-7}$  M CGP 20712A, but was blocked by preincubation with  $10^{-6}$  M nadolol and  $10^{-7}$  M ICI 118,551.

#### Discussion

This study investigated whether the  $\beta_3$ -adrenoceptor agonist, BRL 37344, modulated Na $^+$ ,K $^+$ -pump activity in isolated rat soleus muscle, as well as the mechanisms of its actions. The first main finding was that BRL 37344 stimulates the Na $^+$ ,K $^+$ -pump, and that this occurred via the  $\beta_2$ -adrenoceptors, rather than the  $\beta_3$ -adrenoceptors. The second main finding using the more selective  $\beta_3$ -adrenoceptor agonist, CL 316,243 (Dolan *et al.*, 1994), was that any  $\beta_3$ -adrenoceptors expressed in skeletal muscle

are not involved in  $\mathrm{Na}^+$  and  $\mathrm{K}^+$  homeostasis. Finally, NO did not appear to mediate  $\mathrm{Na}^+$ ,  $\mathrm{K}^+$ -pump stimulation in rat skeletal muscle.



**Figure 6** Effect of the  $β_1/β_2$ -antagonist, nadolol, and the NOS inhibitor, L-NAME, on the BRL 37344-induced force recovery in high [K $^+$ ] $_0$ -depressed rat soleus muscles. Muscles were mounted on force transducers and stimulated at 60 Hz (0.2 ms, 12 V) for 2.0 s every 10 min for 20 min, were then incubated in KR containing high [K $^+$ ] $_0$  (11 mM) and stimulated for a further 170 min. At 90 min of stimulation, the following additions were made; (a)  $10^{-5}$  M BRL 37344;  $10^{-7}$  M BRL 37344 or both  $10^{-5}$  M nadolol and  $10^{-5}$  M BRL 37344, or (b)  $10^{-5}$  M BRL 37344; or both  $10^{-5}$  M L-NAME and  $10^{-5}$  M BRL 37344. (a) Nadolol or (b) L-NAME were added 15 min before the addition of BRL 37344), or are mean  $\pm$  s.e.m. ( $10^{-5}$  M nadolol  $\pm$  10 m BRL 37344) or mean  $\pm$  s.e.m. ( $10^{-5}$  M BRL 37344) or mean  $\pm$  s.e.m. ( $10^{-5}$  M BRL 37344) or clarity; n=4–8.

BRL 37344 stimulated the Na<sup>+</sup>,K<sup>+</sup>-pump via  $\beta_2$ -adrenoceptors The  $\beta_3$ -adrenoceptor agonist, BRL 37344, induced a concentration- and time-dependent reduction in intracellular Na + content that was significant after only 2 min of incubation. As this effect was blocked by ouabain, it was mediated via Na<sup>+</sup>,K<sup>+</sup>-pump stimulation, as also demonstrated by the BRL 37344-induced increase in ouabain-sensitive <sup>86</sup>Rb<sup>+</sup> uptake. The decrease in intracellular Na+ content could not be attributed to inhibition of Na+ influx, the initial rate of <sup>22</sup>Na <sup>+</sup> influx being unaffected by BRL 37344. However, the reduction in intracellular Na+ content induced by BRL 37344 was completely blocked by preincubation with the  $\beta_1/\beta_2$ -adrenoceptor antagonist, nadolol, but not by preincubation with the  $\beta_3$ -adrenoceptor antagonist, SR 59230A  $(10^{-7} \,\mathrm{M})$ . At this concentration, we demonstrated that SR 59230A does not act on the  $\beta_2$ -adrenoceptors since there was no effect of preincubation with this antagonist, on the reduction in intracellular Na+ content induced by the  $\beta_2$ -adrenoceptor agonist, salbutamol (10<sup>-7</sup> M). At the concentrations of  $10^{-6}$  M and  $3 \times 10^{-6}$  M, SR 59203A has previously been shown to act as a selective  $\beta_3$ -adrenoceptor antagonist; having no effect on the increase in cAMP induced with salbutamol in rat cerebellum (Nisoli et al., 1996) and completely blocking the relaxant effects of several selective  $\beta_3$ -adrenoceptor agonists in rat colon (Kaumann and Molenaar, 1996), respectively. However, at the higher concentration of 10<sup>-5</sup> M, SR 59230A appeared to act also on the  $\beta_2$ -adrenoceptors in the present study since the reductions in intracellular Na<sup>+</sup> content induced by BRL 37344 and salbutamol were both slightly attenuated ( $\sim 10-13\%$ , P<0.05) by this preincubation. The lack of selectivity of SR 59230A at a concentration of  $10^{-5}$  M is likely to reflect the considerable heterogeneity that exists between the structure of the  $\beta_1$ -,  $\beta_2$ - and  $\beta_3$ -adrenoceptors, their structure between species, and also between the affinities of these  $\beta$ -adrenoceptors for antagonists and agonists (Arch, 2000). We also demonstrated that nadolol prevents the reduction in intracellular Na<sup>+</sup> content induced by salbutamol. Importantly, salbutamol has previously been shown to be highly selective for the  $\beta_2$ -adrenoceptors (Baker, 2005).

**Table 4** Effect of SR 59230A, nadolol, ICI 118,551, CGP 20712A on the noradrenaline-induced change in intracellular Na<sup>+</sup> content in rat soleus muscle

Preincubation (15 min)	Incubation (30 min)	Intracellular Na $^+$ content ( $\mu$ mol(g wet wt) $^{-1}$ )
No additions	No additions	15.9±0.5
No additions	10 <sup>-5</sup> м Noradrenaline	$7.6\pm0.3*$
10 <sup>-7</sup> м SR 59230A	10 <sup>-7</sup> м SR 59230A	$14.1 \pm 0.3$
10 <sup>-6</sup> м Nadolol	$10^{-6}$ м Nadolol	$14.7 \pm 0.3$
10 <sup>-7</sup> м ICI 118,551	10 <sup>-7</sup> м ICI 118,551	$16.9\pm0.4$
10 <sup>-7</sup> м СGР 20712A	10 <sup>-7</sup> м CGP 20712A	16.0+1.4
10 <sup>-7</sup> м SR 59230A	$10^{-7}$ M SR 59230A + $10^{-5}$ M noradrenaline	6.9 <sup>+</sup> 0.2*
10 <sup>-6</sup> м Nadolol	$10^{-6}$ M Nadolol $+ 10^{-5}$ M noradrenaline	14.7 + 0.3 <sup>†</sup>
10 <sup>-7</sup> м ICI 118,551	$10^{-7}$ M ICI 118,551 + $10^{-5}$ M noradrenaline	15.7 + 0.1 <sup>†</sup>
10 <sup>-7</sup> м СGP 20712A	$10^{-7} \text{ M CGP } 20712 \text{A} + 10^{-5} \text{ M noradrenaline}$	7.7±0.8*

Abbreviations: CGP 20712A, 2-hydroxy-5-(2-[{hydroxy-3-(4-[1-methyl-4-trifluoromethyl-2-imidazolyl]phenoxy)propyl}amino]ethyoxy) benzamide; ICI 118,551, (–)-1-(2,3-[dihydro-7-methyl-1*H*-inden-4-yl]oxy)-3-([1-methylethyl]-amino)-2-butanol; SR 59230A, 3-(2-ethylphenoxy)-1-([1*S*]-1,2,3,4-tetrahydronapth-1-ylamino)-2*S*-2-propranol oxalate.

Muscles were preincubated for 15 min without or with  $10^{-7}$  M SR 59230A,  $10^{-6}$  M nadolol,  $10^{-7}$  M ICI 118,551 or  $10^{-7}$  M CGP 20712A, were incubated for 30 min without or with the indicated additions and then treated as described in Table 1. Data are means  $\pm$  s.e.m., n = 4–13. \*P < 0.001 vs control,  $^{\dagger}P < 0.001$  vs  $10^{-5}$  M noradrenaline.

Furthermore, the force recovery induced by BRL 37344 in high [K<sup>+</sup>]<sub>o</sub>-depressed muscles was completely blocked by preincubation with nadolol. Although nadolol acts as a  $\beta_1/\beta_2$ -adrenoceptor antagonist, the effects shown here with nadolol are mediated via the  $\beta_2$ -adrenoceptors since the reduction in intracellular Na<sup>+</sup> content induced with 10<sup>-7</sup> M BRL 37344 was blocked by preincubation with the selective  $\beta_2$ -antagonist, ICI 118,551 ( $10^{-7}$  M,  $10^{-5}$  M) (O'Donnell and Wanstall, 1980), but not with the selective  $\beta_1$ -adrenoceptor antagonist, CGP 20712A  $(10^{-7} \text{ M}, 10^{-5} \text{ M})$  (Dooley et al., 1986). The selectivity of these antagonists was demonstrated by the suppression of the reduction in intracellular Na<sup>+</sup> content induced with the  $\beta_2$ -adrenoceptor agonist salbutamol  $(10^{-7} \text{ M})$  by preincubation with ICI 118,551  $(10^{-5} \text{ M})$ but not CGP 20712A (10<sup>-5</sup> M). Furthermore, in rat soleus muscle, salbutamol was at least 100 times more potent than the  $\beta_1$ -adrenoceptor agonist H133/22 in stimulating the Na<sup>+</sup>,K<sup>+</sup>-pump (Clausen and Flatman, 1980). Taken together, these results demonstrate that at concentrations exceeding 10<sup>-9</sup> M, BRL 37344 rapidly and potently stimulated the Na+,K+-pump in skeletal muscle, but that this effect was mediated by the  $\beta_2$ -, rather than the  $\beta_3$ - or the  $\beta_1$ -adrenoceptors. Previous studies in rat and mouse skeletal muscle using selective  $\beta_2$ -adrenoceptor antagonists have demonstrated that at low concentrations ( $<10^{-9}$  M), BRL 37344 acts as a selective  $\beta_3$ -adrenoceptor agonist, whereas at higher concentrations ( $>10^{-9}$  M), BRL 37344 also activates the  $\beta_2$ -adrenoceptors (Liu *et al.*, 1996; Board *et al.*, 2000). The present results support these findings since no reduction in intracellular Na<sup>+</sup> content was observed with either 10<sup>-10</sup> M or  $10^{-9}$  M BRL 37344.

No effect of selective  $\beta_3$ -adrenoceptor agonist on Na<sup>+</sup>,K<sup>+</sup>-pump in rat soleus muscle

There was no effect of the more selective  $\beta_3$ -adrenoceptor agonist, CL 316,243, on the intracellular Na $^+$  content in skeletal muscle. This contrasts the reduction in intracellular Na $^+$  content induced by BRL 37344 (>10 $^{-9}$  M) that was mediated via  $\beta_2$ -adrenoceptors, providing evidence that CL 316,243 does not act via the  $\beta_2$ -adrenoceptors. Indeed, it has previously been shown in isolated rat soleus and EDL muscles that the proteolysis induced by CL 316,243 was completely abolished by SR 59230A but not ICI 118,551 (Navegantes *et al.*, 2006).

These results with CL 316,243 are in keeping with our observation of a lack of reduction in intracellular Na<sup>+</sup> content with low concentrations of BRL 37344 ( $10^{-10}$  M,  $10^{-9}$  M), which appear to reflect activation independent of the  $\beta_2$ -adrenoceptors (Liu *et al.*, 1996; Board *et al.*, 2000). Together, these findings suggest (i) that if  $\beta_3$ -adrenoceptors are present in skeletal muscle, they are not involved in Na<sup>+</sup> and K<sup>+</sup> homeostasis in skeletal muscle, (ii) that CL 316,243 and low concentrations of BRL 37344 ( $<10^{-9}$  M) do not affect any  $\beta_3$ -adrenoceptors present in rat soleus muscle, or (iii) that  $\beta_3$ -adrenoceptors are expressed in very low amounts, or not at all, in rat soleus muscle. Despite the above-mentioned study finding evidence for a functional  $\beta_3$ -adrenoceptor in the regulation of proteolysis (Navegantes *et al.*, 2006), it remains unclear whether  $\beta_3$ -adrenoceptors are

indeed expressed in skeletal muscle. Whereas several studies have found evidence for expression of the  $\beta_3$ -adrenoceptor agonist mRNA (Evans et al., 1996) and protein (Sillence et al., 1993) in rat skeletal muscle, others have not (Granneman et al., 1991; McNeel and Mersmann, 1999). In humans,  $\beta_3$ -mRNA was not detected in several types of skeletal muscle, including the soleus, intercostal, posterior tibialis and gastrocnemius muscles (Thomas and Liggett, 1993). However, these results need to be interpreted with caution, as the muscles were taken post mortem from a 43-year-old man who was brain-dead from a massive cerebral vascular haemorrhage. Indeed, the  $\beta_3$ -adrenoceptor protein was identified in two out of three samples of human gastrocnemius muscle using immunohistochemistry and a highaffinity monoclonal antibody for  $\beta_3$  (Mab72c) (Chamberlain et al., 1999). However, in a later study using the same antibody, the  $\beta_3$ -adrenoceptor protein was not detected in any of three samples of human pectoralis muscle (De Matteis et al., 2002). Further work with additional monoclonal antibodies specific for  $\beta_3$  is therefore required to confirm the presence or absence of  $\beta_3$ -adrenoceptors in skeletal muscle. It is also possible that CL 316,243 and low concentrations of BRL 37344 may act via an as yet, uncharacterized  $\beta$ -adrenoceptor in skeletal muscle (Board et al., 2000). As discussed by Board et al. (2000), the existence of such a receptor would support the discrepancy in results pertaining to  $\beta_3$ -adrenoceptor expression in skeletal muscle.

If the present findings represent a lack of  $\beta_3$ -adrenoceptor stimulation of the Na<sup>+</sup>,K<sup>+</sup>-pump in skeletal muscle, this contrasts the stimulatory effect found in rabbit cardiac myocytes (Bundgaard *et al.*, 2006a). This difference may reflect a tissue- or species-specific effect. Indeed, a species-specific effect has been shown for the negative inotropic effect induced by BRL 37344 (Gauthier *et al.*, 1999).

The physiological catecholamines, adrenaline and noradrenaline, activate the Na+,K+-pump in skeletal muscle, leading to increased Na+ efflux and K+ influx and membrane hyperpolarization (Clausen and Flatman, 1977, 1980). As the  $\beta_3$ -adrenoceptor demonstrates a much higher (<30-fold) affinity for noradrenaline than adrenaline, and the opposite is true for the  $\beta_2$ -adrenoceptor in CHO cells (Hoffmann et al., 2004), the noradrenaline-induced Na<sup>+</sup>, K<sup>+</sup>pump activation in skeletal muscle may be mediated via the  $\beta_3$ -adrenoceptors. Indeed, in rabbit cardiac myocytes, the noradrenaline-induced increase in Na+,K+-pump-mediated current was resistant to nadolol (Bundgaard et al., 2006a). However, we found that the reduction in intracellular Na + content induced by noradrenaline (10<sup>-5</sup> M) was blocked by preincubation with nadolol (10<sup>-6</sup> M) and ICI 118,551  $(10^{-7} \text{ M})$ , but not by SR 59230A8  $(10^{-7} \text{ M})$  or CGP 20712A  $(10^{-7} \,\mathrm{M})$ . Thus, in isolated rat soleus muscle, noradrenalineinduced Na<sup>+</sup>,K<sup>+</sup>-pump stimulation occurred via the  $\beta_2$ adrenoceptors, rather than the  $\beta_3$ - or  $\beta_1$ -adrenoceptors.

*No evidence for NO-induced Na*<sup>+</sup>,*K*<sup>+</sup>-*pump stimulation* In skeletal muscle, NO is continuously produced via the enzymatic action of NOS. At rest, NO production is low. However, during periods of repeated action potentials it may increase by up to 100% (Kobzik *et al.*, 1994). Endogenous NO

KT Murphy et al

depresses contractile function, as evidenced by an elevation in submaximal force with the addition of several exogenous NOS inhibitors in rat diaphragm muscle (Kobzik et al., 1994; Reid et al., 1998). Moreover, this effect was reversed with the addition of the exogenous NO donors, S-nitroso-N-acetylcysteine and SNP (Kobzik et al., 1994). In cardiac myocytes, NO signalling is generally restricted to sarcolemmal microdomains (Barouch et al., 2002) that contain significant amounts of Na+,K+-pumps (Liu et al., 2003). Owing to this colocalization, any NO-induced stimulation of the Na<sup>+</sup>,K<sup>+</sup>pump would occur very selectively and rapidly, and therefore, represent a potentially important localized activator of the Na<sup>+</sup>,K<sup>+</sup>-pump. However, in the present study, there was no effect of L-NAME or ODQ, which inhibit NO production and NO-activated guanylyl cyclase, respectively, on the reduction in intracellular Na+ content induced by BRL 37344. There was also no effect of preincubation with L-NAME on the reduction in intracellular Na<sup>+</sup> content induced by salbutamol or on the BRL 37344-induced force recovery in high [K<sup>+</sup>]<sub>o</sub>-depressed muscles. Furthermore, neither of the NO donors, SNP nor SNAP, reduced intracellular Na<sup>+</sup> content. These results therefore indicate that the NO signalling system is not causing stimulation of the Na<sup>+</sup>,K<sup>+</sup>-pump in skeletal muscle. This lack of effect of NO on the Na<sup>+</sup>,K<sup>+</sup>-pump cannot be explained by an absence of NO signalling in skeletal muscle, since a pathway involving NOS, soluble guanylyl cyclase, cGMP and PKG was shown in isolated rat soleus muscle (Young and Leighton, 1998a, b). Conversely, this lack of effect may reflect that NO-induced stimulation of the Na<sup>+</sup>,K<sup>+</sup>-pump is tissue-dependent. In rabbit cardiac myocytes, L-NAME completely abolished the increase in Na<sup>+</sup>,K<sup>+</sup>-pump-mediated current induced by BRL 37344 (Bundgaard et al., 2006a), and Na<sup>+</sup>,K<sup>+</sup>-pump activity was also stimulated by SNP (William et al., 2005). Furthermore, in mouse cardiac vesicles, knockout of the NOS isoforms, nNOS and eNOS, significantly depressed Na<sup>+</sup>,K<sup>+</sup>pump activity (Zhou et al., 2002). In contrast, both SNP and SNAP reduced Na<sup>+</sup>,K<sup>+</sup>-pump activity in the renal medulla of rats (McKee et al., 1994; Beltowski et al., 2003), and SNAP reduced Na<sup>+</sup>,K<sup>+</sup>-pump activity in rat liver cells (Muriel and Sandoval, 2000).

BRL 37344-induced rapid force recovery in muscles depressed by high  $[K^+]_o$ 

In humans, intense exercise has been shown to increase interstitial [K<sup>+</sup>] to as high as 11–12 mM (Nordsborg *et al.*, 2003). Such a large increase reduces the *trans*-sarcolemmal [K<sup>+</sup>] gradient, leading to membrane depolarization and inactivation of the voltage-gated Na<sup>+</sup> channels (Ruff, 1996). This results in loss of muscle excitability, and subsequently, a reduction in force production. Indeed, in the present study, increasing the [K<sup>+</sup>]<sub>o</sub> from 4 to 11 mM reduced force to  $\sim$ 16% of initial force. However, BRL 37344 (10<sup>-5</sup> M) recovered force production to  $\sim$ 92% of the initial level at only 10 min after the onset of exposure. This BRL 37344-induced force recovery is likely to reflect Na<sup>+</sup>,K<sup>+</sup>-pump activation, leading to membrane hyperpolarization and hence, increased force production. This is closely similar to the rapid force recovery induced by  $\beta_2$ -adrenoceptor agonists in high [K<sup>+</sup>]-depressed

muscles as found in this study and earlier (Clausen *et al.*, 1993). In keeping with this, the effect was suppressed by preincubation with nadolol.

#### **Implications**

Several clinical uses of  $\beta_3$ -adrenoceptor agonists have been proposed, including for the treatment of diabetes and obesity (Arch and Wilson, 1996), urinary bladder dysfunction (Badawi et al., 2005), bronchoconstriction (Broadley, 2006) and heart failure (Bundgaard et al., 2006b). Results from the present study suggest that the use of BRL 37344 in these clinical settings could be jeopardized by some of the well-known adverse effects seen clinically when using  $\beta_2$ -adrenoceptor agonists (Broadley, 2006). As BRL 37344 at concentrations exceeding  $10^{-9}\,\mathrm{M}$  appears to act as a  $\beta_2$ -adrenoceptor agonist in skeletal muscle, administration of this compound may induce hypokalemia via Na<sup>+</sup>,K<sup>+</sup>pump stimulation. Indeed, in conscious rabbits, intravenous administration of BRL 37344 led to a significant reduction in plasma K<sup>+</sup> levels, that was even greater than that induced by the administration of salbutamol (Reverte et al., 1993). Thus, clinical use of  $\beta_3$ -adrenoceptor stimulation might require the use of more selective agonists, such as CL 316, 243.

In conclusion, the  $\beta_3$ -adrenoceptor agonist BRL 37344 stimulated the Na<sup>+</sup>,K<sup>+</sup>-pump to reduce intracellular Na<sup>+</sup> content, an effect that was mediated via the  $\beta_2$ -, rather than the  $\beta_3$ -adrenoceptors. In addition, results with the more selective  $\beta_3$ -adrenoceptor agonist CL 316,243 indicated that any  $\beta_3$ -adrenoceptors expressed in rat soleus muscle are unlikely to be involved in Na<sup>+</sup> and K<sup>+</sup> homeostasis in this tissue. Finally, NO did not seem to mediate Na<sup>+</sup>,K<sup>+</sup>-pump stimulation in skeletal muscle.

#### **Acknowledgements**

We thank Ann-Charlotte Andersen, Tove Lindahl Andersen, Vibeke Uhre and Marianne Stürup Johansen for skilled technical assistance. We thank Professor Helge H Rasmussen for proposing experiments with BRL 37344 in skeletal muscle, and also for useful discussions. This study was supported by grants from the Velux and the Lundbeck Foundation.

#### Conflict of interest

The authors state no conflict of interest.

#### References

Arch JR, Ainsworth AT, Ellis RD, Piercy V, Thody VE, Thurlby PL *et al.* (1984). Treatment of obesity with thermogenic  $\beta$ -adrenoceptor agonists: studies on BRL 26830A in rodents. *Int J Obes* 8: 1–11. Arch JR, Kaumann AJ (1993).  $\beta_3$  and atypical  $\beta$ -adrenoceptors. *Med* 

Res Rev 13: 663-729.

Arch JR, Wilson S (1996). Prospects for  $\beta_3$ -adrenoceptor agonists in the treatment of obesity and diabetes. *Int J Obes Relat Metab Disord* **20**: 191–199.

- Arch JRS (2000).  $\beta_3$ -adrenergic ligands. In: Strosberg AD (ed). *The*  $\beta_3$ -adrenergic receptor. Taylor and Francis: London, pp 48–76.
- Badawi JK, Uecelehan H, Hatzinger M, Michel MS, Haferkamp A, Bross S (2005). Relaxant effects of  $\beta$ -adrenergic agonists on porcine and human detrusor muscle. *Acta Physiol Scand* **185**: 151–159.
- Baker JG (2005). The selectivity of *β*-adrenoceptor antagonists at the human  $β_1$ ,  $β_2$  and  $β_3$  adrenoceptors. Br J Pharmacol 144: 317–322.
- Barouch LA, Harrison RW, Skaf MW, Rosas GO, Cappola TP, Kobeissi ZA *et al.* (2002). Nitric oxide regulates the heart by spatial confinement of nitric oxide synthase isoforms. *Nature* **416**: 337-339
- Beltowski J, Marciniak A, Wojcicka G, Gorny D (2003). Nitric oxide decreases renal medullary Na<sup>+</sup>, K<sup>+</sup>-ATPase activity through cyclic GMP-protein kinase G dependent mechanism. *J Physiol Pharmacol* **54**: 191–210.
- Board M, Doyle P, Cawthorne MA (2000). BRL37344, but not CGP12177, stimulates fuel oxidation by soleus muscle *in vitro*. *Eur J Pharmacol* **406**: 33–40.
- Bond RA, Clarke DE (1988). Agonist and antagonist characterization of a putative adrenoceptor with distinct pharmacological properties from the  $\alpha$  and  $\beta$ -subtypes. *Br J Pharmacol* **95**: 723–734.
- Broadley KJ (2006).  $\beta$ -adrenoceptor responses of the airways: for better or worse? *Eur J Pharmacol* **533**: 15–27.
- Buchanan R, Nielsen OB, Clausen T (2002). Excitation- and  $\beta_2$ -agonist-induced activation of the Na<sup>+</sup>-K<sup>+</sup> pump in rat soleus muscle. *J Physiol* **545**: 229–240.
- Bundgaard H, Garcia A, Hamilton EJ, White CN, Rasmussen HH (2006a).  $\beta_3$ -receptors mediate adrenergic stimulation of the sarcolemmal Na $^+$ -K $^+$  pump. *Biophys J* **90**: 556A.
- Bundgaard H, Garcia A, Tinworth KD, Huang Y, Hunyor S, Rasmussen HH (2006b). The haemodynamic effect of BRL-37344 in sheep before and after heart failure. *Biophys J* 90: 558A.
- Chamberlain PD, Jennings KH, Paul F, Cordell J, Berry A, Holmes SD *et al.* (1999). The tissue distribution of the human  $\beta_3$ -adrenoceptor studied using a monoclonal antibody: direct evidence of the  $\beta_3$ -adrenoceptor in human adipose tissue, atrium and skeletal muscle. *Int J Obes Relat Metab Disord* **23**: 1057–1065.
- Clausen T (2003). Na<sup>+</sup>-K<sup>+</sup> pump regulation and skeletal muscle contractility. *Physiol Rev* 83: 1269–1324.
- Clausen T, Andersen SL, Flatman JA (1993). Na<sup>+</sup>-K<sup>+</sup> pump stimulation elicits recovery of contractility in K<sup>+</sup>-paralysed rat muscle. *J Physiol* **472**: 521–536.
- Clausen T, Everts ME, Kjeldsen K (1987). Quantification of the maximum capacity for active sodium-potassium transport in rat skeletal muscle. *J Physiol* 388: 163–181.
- Clausen T, Flatman JA (1977). The effect of catecholamines on Na-K transport and membrane potential in rat soleus muscle. *J Physiol* **270**: 383–414.
- Clausen T, Flatman JA (1980).  $\beta_2$ -adrenoceptors mediate the stimulating effect of adrenaline on active electrogenic Na-K-transport in rat soleus muscle. *Br J Pharmacol* **68**: 749–755.
- Clausen T, Kohn PG (1977). The effect of insulin on the transport of sodium and potassium in rat soleus muscle. *J Physiol* 265: 19-42
- De Matteis R, Arch JR, Petroni ML, Ferrari D, Cinti S, Stock MJ (2002). Immunohistochemical identification of the  $\beta_3$ -adrenoceptor in intact human adipocytes and ventricular myocardium: effect of obesity and treatment with ephedrine and caffeine. *Int J Obes Relat Metab Disord* **26**: 1442–1450.
- Dolan JA, Muenkel HA, Burns MG, Pellegrino SM, Fraser CM, Pietri F *et al.* (1994). *Beta-3* adrenoceptor selectivity of the dioxolane dicarboxylate phenethanolamines. *J Pharmacol Exp Ther* **269**: 1000–1006.
- Dooley DJ, Bittiger H, Reymann NC (1986). CGP 20712 A: a useful tool for quantitating  $\beta_1$  and  $\beta_2$ -adrenoceptors. *Eur J Pharmacol* 130: 137–139.
- Evans BA, Papaioannou M, Bonazzi VR, Summers RJ (1996). Expression of  $\beta_3$ -adrenoceptor mRNA in rat tissues. *Br J Pharmacol* 117: 210–216.
- Everts ME, Clausen T (1992). Activation of the Na-K pump by intracellular Na in rat slow- and fast-twitch muscle. *Acta Physiol Scand* **145**: 353–362.
- Garthwaite J, Southam E, Boulton CL, Nielsen EB, Schmidt K, Mayer B (1995). Potent and selective inhibition of nitric oxide-sensitive

- guanylyl cyclase by 1*H*-[1,2,4]oxadiazolo[4,3-a]quinoxalin-1-one. *Mol Pharmacol* **48**: 184–188.
- Gauthier *C*, Tavernier G, Trochu JN, Leblais V, Laurent K, Langin D *et al.* (1999). Interspecies differences in the cardiac negative inotropic effects of  $\beta_3$ -adrenoceptor agonists. *J Pharmacol Exp Ther* **290**: 687–693.
- Granneman JG, Lahners KN, Chaudhry A (1991). Molecular cloning and expression of the rat  $\beta_3$ -adrenergic receptor. *Mol Pharmacol* **40**: 895–899.
- Hoffmann C, Leitz MR, Oberdorf-Maass S, Lohse MJ, Klotz KN (2004). Comparative pharmacology of human β-adrenergic receptor subtypes-characterization of stably transfected receptors in CHO cells. *Naunyn Schmiedebergs Arch Pharmacol* **369**: 151–159.
- Holm P, Kankaanranta H, Metsa-Ketela T, Moilanen E (1998). Radical releasing properties of nitric oxide donors GEA 3162, SIN-1 and S-nitroso-N-acetylpenicillamine. Eur J Pharmacol 346: 97–102.
- Kaumann AJ, Molenaar P (1996). Differences between the third cardiac β-adrenoceptor and the colonic β<sub>3</sub>-adrenoceptor in the rat. *Br J Pharmacol* **118**: 2085–2098.
- Kobzik L, Reid MB, Bredt DS, Stamler JS (1994). Nitric oxide in skeletal muscle. *Nature* **372**: 546–548.
- Langin D, Portillo MP, Saulnier-Blache JS, Lafontan M (1991). Coexistence of three  $\beta$ -adrenoceptor subtypes in white fat cells of various mammalian species. *Eur J Pharmacol* **199**: 291–301.
- Liu L, Mohammadi K, Aynafshar B, Wang H, Li D, Liu J *et al.* (2003). Role of caveolae in signal-transducing function of cardiac Na<sup>+</sup>/K<sup>+</sup>-ATPase. *Am J Physiol* **284**: C1550–C1560.
- Liu YL, Cawthorne MA, Stock MJ (1996). Biphasic effects of the  $\beta$ -adrenoceptor agonist, BRL 37344, on glucose utilization in rat isolated skeletal muscle. *Br J Pharmacol* 117: 1355–1361.
- McKee M, Scavone C, Nathanson JA (1994). Nitric oxide, cGMP, and hormone regulation of active sodium transport. *Proc Natl Acad Sci USA* 91: 12056–12060.
- McNeel RL, Mersmann HJ (1999). Distribution and quantification of beta<sub>1</sub>-, beta<sub>2</sub>-, and beta<sub>3</sub>-adrenergic receptor subtype transcripts in porcine tissues. *J Anim Sci* 77: 611–621.
- Muriel P, Sandoval G (2000). Nitric oxide and peroxynitrite anion modulate liver plasma membrane fluidity and Na<sup>+</sup>/K<sup>+</sup>-ATPase activity. *Nitric Oxide* 4: 333–342.
- Navegantes LC, Resano NM, Baviera AM, Migliorini RH, Kettelhut IC (2006). CL 316,243, a selective  $\beta_3$ -adrenergic agonist, inhibits protein breakdown in rat skeletal muscle. *Pflügers Arch* **451**: 617–624.
- Nielsen OB, Clausen T (1996). The significance of active Na<sup>+</sup>,K<sup>+</sup> transport in the maintenance of contractility in rat skeletal muscle. *Acta Physiol Scand* **157**: 199–209.
- Nisoli E, Tonello C, Landi M, Carruba MO (1996). Functional studies of the first selective  $\beta_3$ -adrenergic receptor antagonist SR 59230A in rat brown adipocytes. *Mol Pharmacol* **49**: 7–14.
- Nordsborg N, Mohr M, Pedersen LD, Nielsen JJ, Langberg H, Bangsbo J (2003). Muscle interstitial potassium kinetics during intense exhaustive exercise: effect of previous arm exercise. *Am J Physiol* **285**: R143–R148.
- O'Donnell SR, Wanstall JC (1980). Evidence that ICI 118,551 is a potent, highly  $\beta_2$ -selective adrenoceptor antagonist and can be used to characterize  $\beta$ -adrenoceptor populations in tissues. *Life Sci* 27: 671–677.
- Rees DD, Palmer RM, Schulz R, Hodson HF, Moncada S (1990). Characterization of three inhibitors of endothelial nitric oxide synthase *in vitro* and *in vivo*. *Br J Pharmacol* **101**: 746–752.
- Reid MB, Kobzik L, Bredt DS, Stamler JS (1998). Nitric oxide modulates excitation-contraction coupling in the diaphragm. Comp Biochem Physiol A Mol Integr Physiol 119: 211–218.
- Reverte M, Garcia-Barrado MJ, Hernandez-Garcia FJ, Moratinos J (1993). Coexistence of  $\beta_2$  and  $\beta_3$ -adrenoceptors in plasma potassium control in conscious rabbits. *J Auton Pharmacol* **13**: 227–236.
- Ruff RL (1996). Single-channel basis of slow inactivation of Na channels in rat skeletal muscle. Am J Physiol 271: C971–C981.
- Sejersted OM, Sjøgaard G (2000). Dynamics and consequences of potassium shifts in skeletal muscle and heart during exercise. *Physiol Rev* **80**: 1411–1481.
- Sillence MN, Moore NG, Pegg GG, Lindsay DB (1993). Ligand binding properties of putative  $\beta_3$ -adrenoceptors compared in

- brown adipose tissue and in skeletal muscle membranes. Br J Pharmacol 109: 1157–1163.
- Thomas RF, Liggett SB (1993). Lack of  $\beta_3$ -adrenergic receptor mRNA expression in adipose and other metabolic tissues in the adult human. *Mol Pharmacol* **43**: 343–348.
- William M, Vien J, Hamilton E, Garcia A, Bundgaard H, Clarke RJ *et al.* (2005). The nitric oxide donor sodium nitroprusside stimulates the Na<sup>+</sup>-K<sup>+</sup> pump in isolated rabbit cardiac myocytes. *J Physiol* 565: 815–825.
- Yen TT, McKee MM, Stamm NB (1984). Thermogenesis and weight control. *Int J Obes* 8: 65–78.
- Young ME, Leighton B (1998a). Evidence for altered sensitivity of the nitric oxide/cGMP signalling cascade in insulin-resistant skeletal muscle. *Biochem J* **329**: 73–79.
- Young ME, Leighton B (1998b). Fuel oxidation in skeletal muscle is increased by nitric oxide/cGMP evidence for involvement of cGMP-dependent protein kinase. *FEBS Lett* **424**: 79–83.
- Zhou L, Burnett AL, Huang PL, Becker LC, Kuppusamy P, Kass DA *et al.* (2002). Lack of nitric oxide synthase depresses ion transporting enzyme function in cardiac muscle. *Biochem Biophys Res Commun* **294**: 1030–1035.